

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

**PATIENT**  
DISEASE Lung adenocarcinoma  
NAME  
DATE OF BIRTH  
SEX  
MEDICAL RECORD #

**PHYSICIAN**  
ORDERING PHYSICIAN  
MEDICAL FACILITY  
ADDITIONAL RECIPIENT  
MEDICAL FACILITY ID  
PATHOLOGIST

**SPECIMEN**  
SPECIMEN SITE  
SPECIMEN ID  
SPECIMEN TYPE  
DATE OF COLLECTION  
SPECIMEN RECEIVED

**Genomic Signatures**

**Microsatellite status** - MS-Stable  
**Tumor Mutational Burden** - 5 Muts/Mb

**Gene Alterations**

For a complete list of the genes assayed, please refer to the Appendix.

**EGFR** amplification, L858R  
**MDM2** amplification  
**MTAP** loss  
**CDKN2A/B** CDKN2A loss, CDKN2B loss  
**KDR** amplification  
**MUTYH** P391L  
**RBM10** S167fs\*1

7 Disease relevant genes with no reportable alterations: **KRAS, ALK, BRAF, MET, RET, ERBB2, ROS1**

**Report Highlights**

- Targeted therapies with **NCCN categories of evidence** in this tumor type: Afatinib (p. 9), Dacomitinib (p. 10), Erlotinib (p. 10), Gefitinib (p. 11), Osimertinib (p. 12), Cetuximab (p. 13)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 14)
- Variants in select cancer susceptibility genes to consider for possible **follow-up germline testing** in the appropriate clinical context: **MUTYH** P391L (p. 8)

**GENOMIC SIGNATURES**

**Microsatellite status** - MS-Stable  
**Tumor Mutational Burden** - 5 Muts/Mb

**GENE ALTERATIONS**

**EGFR** - amplification, L858R  
10 Trials see p. 14  
**MDM2** - amplification  
5 Trials see p. 16  
**MTAP** - loss  
1 Trial see p. 17

**THERAPY AND CLINICAL TRIAL IMPLICATIONS**

No therapies or clinical trials. see Genomic Signatures section  
No therapies or clinical trials. see Genomic Signatures section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)		THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)	
Afatinib	1	Cetuximab	2A
Dacomitinib	1	Panitumumab	
Erlotinib	1		
Gefitinib	1		
Osimertinib	1		
none		none	
none		none	

NCCN category

**VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING IN SELECT CANCER SUSCEPTIBILITY GENES**

*Findings below have been previously reported as pathogenic germline in the ClinVar genomic database and were detected at an allele frequency of >10%. See appendix for details.*

**MUTYH - P391L** ..... p. 8

This report does not indicate whether variants listed above are germline or somatic in this patient. In the appropriate clinical context, follow-up germline testing would be needed to determine whether a finding is germline or somatic.

**GENE ALTERATIONS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS**

*For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Alterations section.*

<b>CDKN2A/B - CDKN2A loss, CDKN2B loss</b> ..... p. 7	<b>MUTYH - P391L</b> ..... p. 8
<b>KDR - amplification</b> ..... p. 7	<b>RBM10 - S167fs*1</b> ..... p. 8

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved through a centralized EU procedure or a national procedure in an EU Member State. Therapies, including but not limited to the following, have been approved nationally and may not be available in all EU Member States: Tretinoin, Anastrozole, Bicalutamide, Cyproterone, Exemestane, Flutamide, Goserelin, Letrozole, Leuprorelin, Triptorelin.

SAMPLE

ORDERED TEST #

GENOMIC SIGNATURE

## Microsatellite status

RESULT  
MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors<sup>1-3</sup>, including approved therapies nivolumab and pembrolizumab<sup>4</sup>. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,  $p=0.001$ )<sup>5</sup>.

FREQUENCY & PROGNOSIS

MSI-H is generally infrequent in NSCLC, reported in fewer than 1% of samples across several large studies<sup>6-11</sup>, whereas data on the reported incidence of MSI-H in SCLC has been limited and conflicting<sup>12-15</sup>. One study reported MSI-H in lung adenocarcinoma patients with smoking history, and 3 of 4 MSI-H patients examined also had metachronous carcinomas in other organs, although this has not been investigated in large scale studies<sup>6</sup>. Published data investigating the prognostic implications of MSI in NSCLC are limited (PubMed, Oct 2021).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor<sup>16</sup>. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2<sup>16-18</sup>. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers<sup>19-21</sup>. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins<sup>16,18,20-21</sup>.

GENOMIC SIGNATURE

## Tumor Mutational Burden

RESULT  
5 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1<sup>22-24</sup>, anti-PD-1 therapies<sup>22-25</sup>, and combination nivolumab and ipilimumab<sup>26-31</sup>. Multiple clinical trials of PD-1- or PD-L1-targeting immune checkpoint inhibitors or combination of PD-1 and CTLA-4 inhibitors in NSCLC have reported that patients with tumors harboring TMB  $\geq 10$  Muts/Mb derive greater clinical benefit from these therapies than those with TMB  $< 10$  Muts/Mb (based on this assay or others); similarly, higher efficacy of anti-PD-1 or anti-PD-L1 immunotherapy for treatment of patients with NSCLC, compared with the use of chemotherapy, has been observed more significantly in cases of TMB  $\geq 10$  Muts/Mb (based on this assay or others);<sup>22-23,26-28,32-39</sup>. Improved OS of patients with NSCLC treated with pembrolizumab plus chemotherapy relative to chemotherapy only<sup>40</sup>, or those treated with nivolumab plus ipilimumab

also relative to chemotherapy<sup>41</sup>, has been observed across all TMB levels.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that unspecified lung non-small cell lung carcinoma (NSCLC), lung adenocarcinoma, and lung squamous cell carcinoma (SCC) samples harbored median TMBs between 6.3 and 9 Muts/Mb, and 12% to 17% of cases had an elevated TMB of greater than 20 Muts/Mb<sup>42</sup>. Lower TMB is observed more commonly in NSCLCs harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are commonly observed in elevated TMB cases<sup>43</sup>. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC<sup>44-45</sup>, several other large studies did find a strong association with increased TMB<sup>46-49</sup>. TMB  $> 10$  muts/Mb was found to be more frequent in NSCLC metastases compared with primary tumors for both adenocarcinoma (38% vs. 25%) and SCC (41% vs. 35%) subtypes<sup>50</sup>. A meta-analysis of 19 studies of immune checkpoint inhibitor-treated NSCLC ( $n = 2,315$  patients) demonstrated that high TMB predicted a significantly longer OS than low TMB (HR = 0.70), and within the high TMB group, immunotherapy was associated with an improved PFS (HR = 0.62,  $P < 0.001$ ), OS (HR = 0.67,  $P < 0.001$ ) and a higher response rate (OR = 2.35,  $P < 0.001$ ) compared to chemotherapy<sup>51</sup>. In contrast, a large study of Chinese patients with untreated lung

adenocarcinoma reported a shorter median OS for tumors with a higher number of mutations in a limited gene set compared with a lower mutation number (48.4 vs. 61.0 months)<sup>44</sup>. Another study of patients with NSCLC treated with EGFR inhibitors or platinum doublet chemotherapy found elevated TMB to be correlated with poorer prognosis, as well as finding lower TMB in combination with PD-L1 negative status to be significantly associated with longer median survival in patients with lung adenocarcinoma<sup>52</sup>. However, no significant prognostic association of TMB and/or PD-L1 status with survival has been reported in patients with lung SCC<sup>52-53</sup>.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma<sup>54-55</sup> and cigarette smoke in lung cancer<sup>32,56</sup>, treatment with temozolomide-based chemotherapy in glioma<sup>57-58</sup>, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes<sup>59-63</sup>, and microsatellite instability (MSI)<sup>59,62-63</sup>. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents<sup>22-23,26-28,32-39,64</sup>.

ORDERED TEST #

**GENE**  
**EGFR**

**ALTERATION**  
amplification, L858R

**TRANSCRIPT ID**  
NM\_005228

**CODING SEQUENCE EFFECT**  
2573T>G

**VARIANT ALLELE FREQUENCY (% VAF)**  
84.6%

**POTENTIAL TREATMENT STRATEGIES**

**— Targeted Therapies —**

For patients with non-small cell lung cancer, EGFR activating mutations may predict sensitivity to EGFR TKIs, including erlotinib<sup>65</sup>, gefitinib<sup>66</sup>, afatinib<sup>67</sup>, dacomitinib<sup>68</sup>, and osimertinib<sup>69</sup>; however, the data for patients with other tumor types are limited<sup>70-75</sup>. EGFR amplification or expression in patients with non-small cell lung cancer may be associated with benefit from the anti-EGFR antibodies cetuximab<sup>76-77</sup> or necitumumab<sup>78</sup>. Although meta-analyses demonstrate that increased EGFR copy number is significantly associated with improved ORR, PFS, and OS on first-generation EGFR TKIs<sup>79-82</sup>, the magnitude of clinical benefit is limited for patients with EGFR amplification and without sensitizing EGFR mutations when comparing first- or second generation EGFR TKIs to control treatment<sup>83-88</sup>. In the Phase 3 IPASS trial, patients with unmutated, amplified EGFR had a significantly shorter PFS when treated with gefitinib as compared to carboplatin/paclitaxel (HR 3.85; 95% CI, 2.09 to 7.09)<sup>83</sup>. Biomarker analysis of the LUX-Lung 8 trial in squamous NSCLC, which included only a small subset of patients with EGFR mutations (6%), did not observe a significant association of EGFR expression with outcomes on afatinib or erlotinib<sup>89</sup>. A retrospective study in China reported that EGFR amplification was associated with a significantly improved median PFS (5.0 vs

2.0 months) and a similar median OS (16.6. vs. 15.4 months) for patients with unmutated EGFR treated with gefitinib or erlotinib<sup>90</sup>. The Phase 1 CHRYSALIS study of amivantamab monotherapy or in combination with lazertinib for the treatment of EGFR-mutated non-small cell lung cancer (NSCLC) has produced encouraging preliminary results for treatment-naïve patients and patients who relapsed after treatment with osimertinib with and without chemotherapy, including osimertinib-relapsed patients with biomarkers indicating EGFR/MET-based osimertinib resistance<sup>91-94</sup>. In a Phase 1 trial, the HER3-targeted antibody patritumab deruxtecan elicited an ORR of 39% (22/57, 1 CR) and a median PFS of 8.2 months for patients with non-small cell lung cancer previously treated with an EGFR TKI, many of whom displayed TKI resistance alterations<sup>95</sup>. A Phase 1 trial evaluating the EGFR inhibitor AZD3759 reported a reduction in the volume of brain metastases in 40% (8/20) of patients with previously treated non-small cell lung cancer (NSCLC) harboring either the EGFR L858R alteration or EGFR exon 19 deletion, including 3 confirmed PRs and 3 unconfirmed PRs<sup>96-97</sup>. In a Phase 1/2 trial for advanced NSCLC, the brain-penetrant third-generation EGFR TKI lazertinib enabled ORRs of 54% (69/127) for all evaluable patients and 44% (8/18, intracranial) for patients with brain metastases<sup>98</sup>. A Phase 1 trial evaluating the irreversible pan-HER inhibitor FCN-411 for NSCLC patients who had EGFR mutations and experienced disease progression on standard treatments reported an ORR of 15% with 10/67 patients achieving PR, and a DCR of 73% with 39 additional patients achieving SD<sup>99</sup>. OR was observed in a numerically higher proportion of patients with the EGFR T790M mutation than those without this mutation<sup>99</sup>.

**— Nontargeted Approaches —**

Patients with EGFR-mutated non-squamous metastatic non-small cell lung cancer previously treated with EGFR TKI have benefited from immune checkpoint inhibitors combined with

anti-angiogenic and chemotherapy, particularly atezolizumab plus bevacizumab plus carboplatin and paclitaxel (OS HR 0.61 compared with bevacizumab/chemotherapy)<sup>100-102</sup> or sintilimab plus bevacizumab biosimilar plus cisplatin and pemetrexed (PFS HR 0.46 compared with chemotherapy alone)<sup>103</sup>.

**FREQUENCY & PROGNOSIS**

EGFR mutation has been reported in 12-36% of lung adenocarcinomas<sup>48,104-105</sup> and in 4% of lung squamous cell carcinomas<sup>106</sup>. Amplification of EGFR has been variously reported in 4-42% of non-small cell lung carcinoma (NSCLC) samples<sup>105-109</sup>. EGFR protein expression/overexpression has been reported in up to 70% of NSCLC cases<sup>107-112</sup>. In addition, expression of EGFR protein has been shown to be higher in lung squamous cell carcinoma samples as compared to lung adenocarcinoma<sup>113-114</sup>. In patients with lung adenocarcinoma, EGFR gene amplification was a predictor of poor disease-free survival<sup>115</sup>. In patients with lung adenocarcinoma, EGFR mutation was a predictor of poor overall survival<sup>115-116</sup>. However, EGFR mutations have been reported to predict improved survival in patients with resected Stage 1-3 lung adenocarcinoma<sup>117</sup> or resected Stage 1 NSCLC<sup>118</sup>.

**FINDING SUMMARY**

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide<sup>119</sup>. Amplification of EGFR has been associated with increased expression of EGFR mRNA and protein in several cancer types<sup>108,120-121</sup>. EGFR L858R is located in the kinase domain and is encoded by exon 21. EGFR L858R has been characterized as activating<sup>122-124</sup> and patients with the L858R mutation have been shown to be sensitive to EGFR tyrosine kinase inhibitors, such as erlotinib, gefitinib<sup>122-124</sup>, and afatinib<sup>125</sup>.

ORDERED TEST #

GENE  
**MDM2**

ALTERATION  
amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

MDM2 antagonists disrupt the MDM2-p53 interaction, thereby stabilizing p53<sup>126</sup>. Preclinical studies have suggested that the amplification of MDM2, in the absence of concurrent TP53 mutations, may increase sensitivity to these agents<sup>127-128</sup>. Preliminary Phase 1 studies of the MDM2-p53 antagonist alrizomadlin (APG-115) reported a PR in a patient with liposarcoma harboring an MDM2 amplification and wildtype for TP53 and SD in 21%-38% (6/28 and 5/13, respectively) of patients in genomically unselected solid tumors<sup>129-130</sup>. A Phase 2 trial of alrizomadlin in combination with pembrolizumab reported a PR in 1 of 3 patients with malignant peripheral nerve sheath tumor that had failed standard therapy, as well as PRs in patients with multiple types of solid tumors that had failed immunotherapy, including 1 out of 14 patients with non-small cell lung cancer; 1 out of 5 patients

with urothelial carcinoma; and 2 out of 5, 1 out of 5, and 1 out of 11 patients with mucosal, uveal, and cutaneous melanoma, respectively<sup>131</sup>. Phase 1b studies of the MDM2 inhibitor idasanutlin for refractory AML in combination with cytarabine or venetoclax reported anti-leukemic response rates of 33% (25/75) and 37% (11/30), respectively<sup>132-133</sup>; clinical benefit (58% ORR, 7/12) with idasanutlin monotherapy has been reported for patients with polycythemia vera<sup>134</sup>. The dual MDM2/MDM4 inhibitor ALRN-6924 led to an ORR of 27% (4/15) for patients with TP53 wildtype peripheral T-cell lymphoma in a Phase 2 study<sup>135</sup>; responses have also been observed in TP53 wildtype AML, MDS, Merkel cell carcinoma, colorectal cancer, and liposarcoma<sup>136-137</sup>.

FREQUENCY & PROGNOSIS

In the TCGA datasets, amplification of MDM2 has been reported in 8% of lung adenocarcinoma cases<sup>105</sup> and 2% of lung squamous cell carcinoma cases<sup>106</sup>. Separate studies have reported MDM2 amplification at similar incidences of 6-7% in non-small cell lung cancer (NSCLC), mainly in patients with adenocarcinoma, but a higher incidence of 21% (24/116) has also been observed, with amplification found in various NSCLC subtypes<sup>138-140</sup>. The role of MDM2 expression/amplification as a prognostic marker is complex,

with some studies showing a negative and others a positive effect on survival in patients with NSCLC<sup>138,140-142</sup>.

FINDING SUMMARY

MDM2 encodes an E3 ubiquitin protein ligase, which mediates the ubiquitination and subsequent degradation of p53, Rb1, and other proteins<sup>143-145</sup>. MDM2 acts to prevent the activity of the tumor suppressor p53; therefore, overexpression or amplification of MDM2 may be oncogenic<sup>146-147</sup>. Overexpression or amplification of MDM2 is frequent in cancer<sup>148</sup>. Although two retrospective clinical studies suggest that MDM2 amplification may predict a short time-to-treatment failure on anti-PD-1/PD-L1 immune checkpoint inhibitors, with 4/5 patients with MDM2 amplification<sup>149</sup> and 2/3 patients with MDM2 or MDM4 amplification<sup>150</sup> experiencing tumor hyperprogression, amplification of MDM2 or MDM4 was not associated with shorter progression-free survival (PFS) in a retrospective analysis of non-small cell lung cancer (NSCLC) outcomes with immune checkpoint inhibitors (hazard ratio of 1.4, p=0.44)<sup>36</sup>. The latter study reported PFS of >2 months for 5/8 patients with MDM2/MDM4 amplification<sup>36</sup>.

ORDERED TEST #

GENOMIC FINDINGS

**GENE**  
**MTAP**

**ALTERATION**  
loss

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

MTAP inactivation produces specific metabolic vulnerabilities that may be sensitive to MAT2A<sup>151-152</sup> or PRMT5 inhibition<sup>152-154</sup>. A Phase 1 trial of MAT2A inhibitor AG-270 reported 1 PR and 2 SDs lasting longer than 6 months for patients with advanced solid tumors displaying MTAP loss<sup>155</sup>. Preclinical data suggest that MTAP loss sensitizes cells to S-adenosyl-L-methionine (SAM)-competitive PRMT5 inhibitors<sup>156</sup>, dual PRMT1 and PRMT5 inhibitors<sup>157-159</sup>, and PRMT5 inhibitors that selectively bind the PRMT5 when complexed with S-methyl-5'-thioadenosine (MTA), such as MRTX1719<sup>160</sup>. In preclinical models, MTAP inactivation showed increased sensitivity

to inhibitors of purine synthesis or purine analogs, especially upon addition of exogenous MTA<sup>161-171</sup>. A Phase 2 study of L-alanosine, an inhibitor of adenine synthesis, as a monotherapy for 65 patients with MTAP-deficient cancers reported no responses and SD for 24% (13/55) of patients<sup>172</sup>. Preclinical and limited clinical evidence suggest MTAP deficiency may confer sensitivity to pemetrexed<sup>173</sup>.

**FREQUENCY & PROGNOSIS**

MTAP loss/homozygous deletion as well as loss of expression has been reported in a wide variety of solid tumors and hematologic cancers<sup>174-175</sup>; such events have been correlated with poor prognosis in a variety of cancer types, including hepatocellular carcinoma<sup>176</sup>, gastrointestinal stromal tumors<sup>177</sup>, mantle cell lymphoma (MCL)<sup>178</sup>, melanoma<sup>179-180</sup>, gastric cancer<sup>181</sup>, myxofibrosarcoma<sup>182</sup>, nasopharyngeal carcinoma<sup>183</sup>, ovarian carcinoma<sup>174</sup> and non-small cell lung cancer<sup>184</sup>. MTAP loss was not prognostic in pediatric B-cell acute lymphocytic leukemia<sup>185</sup> or in astrocytoma<sup>186</sup>. However, MTAP has also

been reported to be overexpressed in colorectal cancer (CRC) samples<sup>187</sup>, and MTAP retention is thought to be important for prostate cancer growth due to continuous supply of SAM<sup>188</sup>. Germline SNPs in MTAP have been correlated with the development of cutaneous melanoma<sup>189-190</sup>, esophageal cancer<sup>191-192</sup>, osteosarcoma<sup>193</sup>, and CRC<sup>194</sup>.

**FINDING SUMMARY**

MTAP encodes S-methyl-5'-thioadenosine (MTA) phosphorylase, a tumor suppressor involved in polyamine metabolism and methionine synthesis, although its enzymatic function is dispensable for its tumor suppressor activity<sup>195-196</sup>. Decreased expression of MTAP leads to MTA accumulation within tumor cells and their microenvironment<sup>176,197-198</sup>, thereby reducing intracellular arginine methylation<sup>152-154</sup> and altering cell signaling<sup>198-199</sup>. MTAP is located at 9p21, adjacent to CDKN2A and CDKN2B, with which it is frequently co-deleted in various cancers. Other alterations in MTAP are rare and have not been extensively characterized.

ORDERED TEST #

**GENE**  
**CDKN2A/B**

**ALTERATION**  
CDKN2A loss, CDKN2B loss

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

Although case studies have reported that patients with breast cancer or uterine leiomyosarcoma harboring CDKN2A loss responded to palbociclib treatment<sup>200-201</sup>, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents<sup>202-208</sup>; it is not known whether CDK4/6 inhibitors would be beneficial in this case. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors<sup>128,209</sup>, the clinical relevance of p14ARF as a predictive biomarker is not clear. Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib<sup>210-213</sup>. There are no drugs that directly target the mutation or loss of CDKN2B in cancer. Because the p15INK4b protein encoded by CDKN2B is known to inhibit CDK4, tumors with CDKN2B mutation or loss may predict sensitivity to CDK4/6 inhibitors, such as ribociclib,

abemaciclib, and palbociclib<sup>203,205-206,214-216</sup>.

**FREQUENCY & PROGNOSIS**

CDKN2A/B loss and CDKN2A mutation have been reported in approximately 19% and 4% of lung adenocarcinomas, respectively<sup>105</sup>. CDKN2A/B loss and CDKN2A mutation have been reported in 26% and 17% of lung squamous cell carcinoma (SCC) samples analyzed in the TCGA dataset, respectively<sup>106</sup>. Loss of p16INK4a protein expression, through CDKN2A mutation, homozygous deletion, or promoter methylation, has been described in 49-68% of non-small cell lung cancer (NSCLC) samples, whereas low p14ARF protein expression has been detected in 21-72% of NSCLC samples<sup>106,217-222</sup>. In patients with lung SCC, loss of CDKN2B associated with poor survival in one study<sup>223</sup>. Loss of p16INK4a protein as well as CDKN2A promoter hypermethylation correlate with poor survival in patients with NSCLC<sup>219,224-226</sup>.

**FINDING SUMMARY**

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b<sup>227-228</sup>. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to

dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control<sup>218,229</sup>. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition<sup>230-231</sup>. One or more alterations observed here are predicted to result in p16INK4a loss of function<sup>232-253</sup>. One or more alterations seen here are predicted to result in p14ARF loss of function<sup>236,253-256</sup>. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b<sup>257</sup>.

**POTENTIAL GERMLINE IMPLICATIONS**

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer<sup>258</sup>. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma<sup>259-260</sup>. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases<sup>261-263</sup>. CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors<sup>264-266</sup>. In the appropriate clinical context, germline testing of CDKN2A is recommended.

**GENE**  
**KDR**

**ALTERATION**  
amplification

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

On the basis of clinical benefit for patients with ccRCC<sup>267-271</sup> and a patient with breast angiosarcoma<sup>272</sup>, high VEGFR-2 expression has been associated with sensitivity to sunitinib. However, because data supporting concordance between VEGFR-2 expression and KDR genomic biomarkers are limited, it is unclear whether these

therapeutic strategies would be beneficial in this case. On the basis of extensive clinical evidence across multiple tumor types, expression of plasma or tumor VEGFR-1 or VEGFR-2 has not been established as a reliable biomarker to predict response to the VEGFA-targeted agent bevacizumab<sup>273-292</sup>.

**FREQUENCY & PROGNOSIS**

Amplification of KDR has been reported in 1% of lung adenocarcinomas<sup>105</sup> and in 5% of lung squamous cell carcinomas<sup>106</sup>. Amplification of chromosome 4q12, including the genes that encode the tyrosine kinases PDGFR-alpha, KIT, and VEGFR-2 (KDR), has been observed in a subset of tumors<sup>293-294</sup>, including 3.5% of lung adenocarcinomas<sup>295</sup>. KDR expression and/or

amplification in patients with non-small cell lung cancer (NSCLC) has been correlated with worse prognosis, angiogenesis, and resistance to platinum chemotherapy<sup>296-297</sup>.

**FINDING SUMMARY**

KDR encodes vascular endothelial growth factor receptor 2 (VEGFR2), a member of the vascular endothelial growth factor receptor (VEGFR) family. It is a receptor tyrosine kinase that transmits signals from VEGFA and is involved in both tumor angiogenesis and vasculogenesis during development<sup>298</sup>. KDR amplification has been reported in many tumor types and may be oncogenic<sup>298</sup>.

ORDERED TEST #

**GENE**  
**MUTYH**

**ALTERATION**

P391L

**TRANSCRIPT ID**

NM\_001048171

**CODING SEQUENCE EFFECT**

1172C>T

**VARIANT ALLELE FREQUENCY (% VAF)**

48.7%

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

There are no therapies or clinical trials available to address MUTYH alterations in cancer.

**FREQUENCY & PROGNOSIS**

In general, somatic MUTYH mutations are infrequently reported across cancer types (COSMIC, 2022)<sup>299</sup>. Monoallelic MUTYH mutation

occurs in 1-2% of the general population<sup>300-301</sup>. There is conflicting data regarding the impact of monoallelic mutations on the risk of developing CRC<sup>302-304</sup>. Patients with MUTYH-mutant CRC were reported to have significantly improved overall survival compared to patients without MUTYH mutation<sup>305</sup>.

**FINDING SUMMARY**

MUTYH (also known as MYH) encodes an enzyme involved in DNA base excision repair, and loss of function mutations in MUTYH result in increased rates of mutagenesis and promotion of tumorigenesis<sup>306</sup>. The two most frequently reported MUTYH loss of function mutations are G382D (also referred to as G396D) and Y165C (also referred to as Y179C)<sup>300-301,307-309</sup>. Numerous other MUTYH mutations have also been shown to result in loss of function<sup>307-310</sup>.

**POTENTIAL GERMLINE IMPLICATIONS**

One or more of the MUTYH variants observed here has been described in the ClinVar database as

a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with MUTYH-associated polyposis (ClinVar, Mar 2022)<sup>311</sup>. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline biallelic MUTYH mutation causes MUTYH-associated polyposis (also known as MYH-associated polyposis or MAP), an autosomal recessive condition characterized by multiple colorectal adenomas and increased lifetime risk of colorectal cancer (CRC)<sup>300,312-314</sup>. MAP accounts for approximately 0.7% of all CRC cases and 2% of early-onset CRC cases<sup>300</sup>. In contrast to CRC, the role of MUTYH mutation in the context of other cancer types is not well established<sup>315-319</sup>. Estimates for the prevalence of MAP in the general population range from 1:5,000-1:10,000<sup>301</sup>. Therefore, in the appropriate clinical context, germline testing of MUTYH is recommended.

**GENE**  
**RBM10**

**ALTERATION**

S167fs\*1

**TRANSCRIPT ID**

NM\_005676

**CODING SEQUENCE EFFECT**

500delC

**VARIANT ALLELE FREQUENCY (% VAF)**

19.0%

**POTENTIAL TREATMENT STRATEGIES**

— Targeted Therapies —

There are no targeted therapies approved or

clinical trials that directly address genomic alterations in RBM10.

**FREQUENCY & PROGNOSIS**

Mutations in RBM10 have been identified in numerous cancers at low frequencies, including non-small cell lung cancer (NSCLC) (7.4%), bladder cancer (5.4%), and thyroid cancer (4.6%)<sup>320-321</sup>. Several reports have analyzed the prognostic relevance of RBM10 mutations and expression levels in cancer, but results have been mixed. RBM10 mutations were associated with shorter OS in lung adenocarcinomas<sup>322-323</sup>, but with enhanced survival in pancreatic ductal adenocarcinomas compared with RBM10 wildtype tumors<sup>324</sup>. Low RBM10 expression was associated with poor prognosis and shorter OS in patients

with hepatocellular carcinoma (HCC) and pancreatic ductal cancer<sup>325-326</sup>, while in lung adenocarcinoma, it was associated with a better prognosis and longer OS<sup>327</sup>.

**FINDING SUMMARY**

RBM10 encodes RNA binding motif protein 10, a nuclear RNA-binding protein involved in the regulation of alternative splicing<sup>328-329</sup>. Germline mutations in RBM10 cause TARP syndrome, an X-linked recessive disorder characterized by development of micrognathia, glossoptosis, and cleft palate<sup>330-331</sup>.



THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

# Afatinib

Assay findings association

**EGFR**  
amplification, L858R

### AREAS OF THERAPEUTIC USE

Afatinib is an irreversible kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4. It is available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) and activating EGFR mutations and for the treatment of patients with advanced squamous NSCLC after progression on platinum-based chemotherapy. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer<sup>67-68,332-333</sup>, whereas data for patients with other tumor types are limited<sup>70-75,334</sup>.

### SUPPORTING DATA

Afatinib enabled 1 PR and 1 SD for 2 patients with EGFR-amplified NSCLC in a Phase 2 study<sup>85</sup>. Afatinib has shown significant clinical activity for patients with NSCLC and the EGFR common sensitizing mutations L858R or exon 19 deletions, based on extensive clinical evidence<sup>67,332,335-338</sup>. Two randomized Phase 3 trials reported significantly improved median PFS from afatinib compared with chemotherapy for patients with EGFR common sensitizing mutations (LUX-Lung 3, 13.6 vs. 6.9 months, HR 0.47, p<0.001; LUX-Lung 6, 11.0 vs. 5.6 months, HR 0.28, p<0.0001)<sup>67,332</sup>. However, while afatinib significantly increased OS relative to chemotherapy for patients with EGFR exon 19 alterations in these two trials (LUX-Lung 3, 33.3 vs. 21.1 months, HR=0.54; LUX-Lung 6, 31.4 vs. 18.4 months, HR=0.64), no significant OS differences were observed in treatment for patients with L858R mutation<sup>125</sup>. A similar alteration-specific difference was observed for EGFR-mutated treatment-naive NSCLC in a retrospective analysis, which reported numerically longer median OS from second- versus first-generation EGFR TKIs (48.8 vs. 26.4 months, HR=0.59) for patients with exon 19 deletions, but no substantial difference for patients with L858R (25.4 vs. 20.6 months, HR=0.90)<sup>335</sup>. A Phase 2b study of first-line afatinib compared with gefitinib, also for NSCLC with exon 19 deletions or L858R, reported similar median OS for the two therapies (27.9 vs. 24.5 months, HR=0.86) but significantly longer time-to-treatment-failure (13.7 vs. 11.5 months, HR=0.75)

and higher ORR (73% vs. 56%, p=0.0018) with afatinib<sup>336</sup>. Patients with metastatic NSCLC and common EGFR mutations who progressed on prior chemotherapy experienced an ORR of 50.0% (30/60) from afatinib in a Phase 4 trial<sup>337</sup>. As first-line therapy for NSCLC with EGFR exon 19 deletions or L858R, prospective or randomized Phase 2 trials have reported a median PFS of 10.2 months and OS of 24.8 months for patients unfit for chemotherapy<sup>338</sup> and an ORR of 72.5% (n=40, 1 CR), DCR of 100% (40/40), and median PFS and OS of 15.2 and 30.0 months, respectively, for elderly patients ≥70 years old<sup>339</sup>. A retrospective study of afatinib administered to Asian patients with NSCLC, 99% of whom were previously treated with erlotinib and/or gefitinib, reported an ORR of 27.4% (63/230) for patients with common sensitizing EGFR mutations and an ORR of 24.4% (105/431) for the entire cohort<sup>340</sup>. In a case report, a patient with NSCLC with exon 19 deletion and leptomeningeal metastases experienced an ongoing 16-month PR from afatinib in extracranial, brain, and leptomeningeal lesions<sup>341</sup>. For patients with erlotinib- or gefitinib-resistant NSCLC and EGFR mutations, Phase 2/3 studies of afatinib treatment have generally reported ORRs of only 7 to 9%<sup>85,342-346</sup>; however, DCRs of more than 50% have been observed<sup>85</sup>. In a Phase 1b or observational study, patients with EGFR-mutated NSCLC who progressed on afatinib experienced further clinical benefit from subsequent treatment with afatinib and cetuximab<sup>347</sup> or osimertinib<sup>348</sup>, respectively. Extensive clinical data have demonstrated that afatinib is effective for patients with EGFR-mutated advanced NSCLC, including exon 19 deletions and L858R mutations, as well as uncommon sensitizing mutations in exons 18 or 20<sup>67,125,332,336,338,340,349</sup>. Afatinib has also shown activity for patients with advanced NSCLC and ERBB2 mutations, most of which were exon 20 insertions<sup>85,350-360</sup>. The randomized Phase 3 LUX-Lung 8 trial comparing afatinib with erlotinib as second-line therapy for advanced lung squamous cell carcinoma (SCC) reported significantly longer median OS (7.9 vs. 6.8 months, HR=0.81), significantly longer median PFS (2.6 vs. 1.9 months, HR=0.81), and higher DCR (51% vs. 40%, p=0.002) for patients treated with afatinib<sup>349</sup>. For patients who progressed on afatinib monotherapy, additional clinical benefit has been reported from afatinib combined with paclitaxel<sup>361</sup>.

THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

## Dacomitinib

Assay findings association

**EGFR**  
amplification, L858R

### AREAS OF THERAPEUTIC USE

Dacomitinib is a second-generation irreversible tyrosine kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4/HER4. It is available in the EU for first-line treatment of patients with advanced non-small cell lung cancer (NSCLC) with EGFR activating mutations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer<sup>67-68,332-333</sup>, whereas data for patients with other tumor types are limited<sup>70-75,334</sup>. Patients with untreated advanced NSCLC and EGFR L858R mutations achieved an ORR of 73% (68/93)<sup>362</sup> and a median OS of 32.5 months with dacomitinib<sup>68</sup>.

### SUPPORTING DATA

A randomized Phase 3 trial in patients with NSCLC with activating EGFR mutations (primarily L858R or exon 19 deletions) reported improved clinical benefit with first-line dacomitinib compared with gefitinib (median OS,

34.1 vs. 26.8 months, HR=0.760; median PFS, 14.7 vs. 9.2 months, HR=0.59)<sup>362-363</sup>; median OS was 34.1 to 36.7 months and ORR was 74.9% to 79.3%, depending on the dosing regimen<sup>364</sup>. A pooled subgroup analysis of patients with NSCLC with activating EGFR mutations reported improved clinical efficacy with dacomitinib treatment compared with erlotinib (median PFS, 14.6 vs. 9.6 months, HR=0.717; median OS, 26.6 vs. 23.2 months, HR=0.737)<sup>365</sup>. Reduced efficacy of dacomitinib treatment in patients with NSCLC harboring the EGFR T790M mutation has been reported in multiple studies<sup>366-368</sup>. A Phase 1 trial of combination dacomitinib and a MEK1/2 inhibitor for patients with KRAS-mutated CRC, NSCLC, or pancreatic cancer reported 20/36 SDs and 16 PDs, however toxicity from this combination prevented long-term treatment in this patient population<sup>369</sup>. A Phase 2 study of dacomitinib in patients with NSCLC who had been previously treated with chemotherapy or erlotinib and were not selected for EGFR mutations reported an ORR of 4.5% (3/66)<sup>367</sup>. In one study, the combination of dacomitinib and crizotinib was ineffective and associated with high toxicity in patients with NSCLC<sup>370</sup>.

## Erlotinib

Assay findings association

**EGFR**  
amplification, L858R

### AREAS OF THERAPEUTIC USE

Erlotinib is an EGFR tyrosine kinase inhibitor. It is available in the EU to treat advanced non-small cell lung cancer (NSCLC) as first-line therapy or switch maintenance therapy for patients with EGFR-activating mutations and as second-line therapy for patients who have progressed on prior chemotherapy. Erlotinib is also available in combination with gemcitabine to treat metastatic pancreatic cancer. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression<sup>65,371-373</sup>. For patients with esophageal or biliary cancer treated with erlotinib or gefitinib, elevated EGFR copy number or amplification is associated with clinical responses and longer survival<sup>374-378</sup>.

### SUPPORTING DATA

For patients with EGFR-mutated non-small cell lung cancer (NSCLC), the Phase 3 EURTAC trial improved PFS with first-line erlotinib relative to platinum-based chemotherapy (9.7 vs. 5.2 months, HR=0.37), though OS

was not prolonged (22.9 vs 19.6 months, HR=0.92)<sup>65,379</sup>. This study and meta-analyses attribute the lack of OS benefit to the effectiveness of post-progression salvage therapy in the control arm<sup>380</sup>. A Phase 3 study reported similar efficacy of erlotinib and gefitinib for patients with EGFR-mutated NSCLC<sup>381</sup>. Patients with EGFR-mutated NSCLC have experienced PFS benefit with the addition of bevacizumab to erlotinib in the first-line setting in Phase 3 trials including the ARTEMIS-CTONG1509 trial for Chinese patients (17.9 vs. 11.2 months, HR=0.55)<sup>382</sup>, the NEJo26 trial for Japanese patients (16.9 vs. 13.3 months, HR=0.605)<sup>383-384</sup>, and the international BEVERLY trial (15.4 vs. 9.7 months, HR=0.60)<sup>385</sup>; OS benefit has not been observed across these studies. In the maintenance setting, Phase 3 trials have reported significantly improved PFS with maintenance erlotinib following first-line platinum-based chemotherapy, with the largest benefit for patients with EGFR mutations<sup>371,386</sup>. In the neoadjuvant setting, a Phase 2 trial reported a numerically improved ORR and significantly longer PFS with erlotinib compared with chemotherapy for patients with EGFR-mutated advanced NSCLC<sup>372</sup>. In the placebo-controlled Phase 3 RELAY trial, the addition of ramucirumab to erlotinib improved PFS for previously untreated patients with NSCLC harboring EGFR L858R or exon 19 deletion (19.4 vs. 12.4 months, HR=0.59)<sup>387</sup>.

THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

# Gefitinib

Assay findings association

**EGFR**  
amplification, L858R

**AREAS OF THERAPEUTIC USE**

Gefitinib is an EGFR tyrosine kinase inhibitor available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) with activating EGFR mutations. Please see the drug label for full prescribing information.

**GENE ASSOCIATION**

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy<sup>373,388-393</sup>, and responses have been reported for patients with EGFR-rearranged NSCLC<sup>394-395</sup>. For patients with esophageal or biliary cancer treated with erlotinib or gefitinib, elevated EGFR copy number or amplification is associated with clinical responses and longer survival<sup>374-378</sup>. Patients with refractory advanced esophageal carcinoma and EGFR amplification derived significant overall survival benefit from gefitinib compared to placebo (HR = 0.21)<sup>374,396</sup>.

**SUPPORTING DATA**

Gefitinib achieved an ORR of 69.8% and OS of 19.2

months as first-line treatment for Caucasian patients with non-small cell lung cancer (NSCLC) and EGFR sensitizing mutations<sup>66</sup>. Phase 3 studies for Japanese patients<sup>390,397</sup> and East Asian patients<sup>83,391</sup> with EGFR-mutated NSCLC reported longer PFS but not longer OS on first-line gefitinib compared with cisplatin and docetaxel or carboplatin and paclitaxel. Retrospective analysis of East Asian patients receiving first-line gefitinib reported greatest PFS benefit among patients with EGFR exon 19 insertions or deletions and shortest PFS for those with exon 20 insertions (1.2 months)<sup>398</sup>. Two Phase 3 trials of the combination gefitinib plus pemetrexed and carboplatin compared with gefitinib alone for patients with advanced NSCLC harboring EGFR activating mutations reported significantly higher ORRs (75.3% and 84% vs. 62.5% and 67%), longer median PFS (16 and 20.9 months vs. 8 and 11.9 months), and longer median OS (50.9 months and not reached vs. 17 and 38.8 months) with combination treatment; however, combination treatment was associated with increased Grade 3 or higher adverse events<sup>399-400</sup>. In a Phase 1 study for treatment-naïve patients with NSCLC, 63% (19/30) of patients experienced PR from the combination of gefitinib and the PD-L1 inhibitor durvalumab<sup>401</sup>.

SAMPLE

THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

# Osimertinib

Assay findings association

## EGFR

amplification, L858R

### AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR tyrosine kinase inhibitor (TKI) that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is available in the EU in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors harbor EGFR T790M mutations or activating mutations, including EGFR exon 19 deletions and exon 21 L858R mutations. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer<sup>69,394,402-404</sup>. Patients with untreated advanced NSCLC and EGFR exon 19 deletions or L858R mutations achieved an ORR of 80% and a median PFS of 21.4 and 14.4 months, respectively<sup>402</sup>.

### SUPPORTING DATA

The Phase 3 FLAURA study reported that, relative to erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (18.9 vs. 10.2 months; HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80) for patients with advanced NSCLC and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858R)<sup>402,405</sup>. In the Phase 3 ADAURA study, patients with early Stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer PFSs on osimertinib compared to placebo in the adjuvant setting (not reached vs. 28.1

months; HR=0.21)<sup>406</sup>. A Phase 1 study reported that T790M-negative patients with acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months<sup>69</sup>. A Phase 1b/2 study evaluating osimertinib in combination with the CD73 inhibitor olectumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/19), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 2 trial of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with untreated advanced non-small cell lung cancer (NSCLC) harboring EGFR del19 or L858R reported no difference in ORR (82% vs 86%) and median PFS (22.1 vs 20.2 months, HR 0.862 p=0.213)<sup>407</sup>. The Phase 2 BOOSTER study of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with advanced NSCLC with EGFR-sensitizing mutations (exon 19 del or L858R) and L790M at progression on prior EGFR TKI reported no difference in ORR (55% vs 55%), median OS (24.0 vs 24.3 months, HR 1.03 p=0.91), or median PFS (15.4 vs 12.3 months, HR 0.96 p=0.83), although improved PFS was observed for the combination in the subgroup of current or former smokers (16.5 vs 8.4, HR 0.52) while nonsmokers had no benefit (HR 1.47)<sup>408</sup>. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively<sup>409</sup>.

SAMPLE

THERAPIES WITH CLINICAL BENEFIT | IN OTHER TUMOR TYPE

ORDERED TEST #

## Cetuximab

Assay findings association

**EGFR**  
amplification, L858R

### AREAS OF THERAPEUTIC USE

Cetuximab is a monoclonal antibody that targets EGFR. It is available in the EU to treat EGFR-expressing RAS wild-type metastatic colorectal cancer (CRC) as monotherapy or combined with chemotherapy. Cetuximab is also available to treat advanced head and neck squamous cell carcinoma (HNSCC) in combination with other therapies. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased survival was not seen in populations that received first-line treatment with EGFR antibodies<sup>410</sup>.

### SUPPORTING DATA

The Phase 3 FLEX study for patients with high EGFR expression in non-small cell lung cancer (NSCLC) demonstrated that treatment with cetuximab plus chemotherapy resulted in longer overall survival compared to chemotherapy alone (12 months vs 9.6 months)<sup>76</sup>. A Phase 2 study of 31 patients with NSCLC

found that the addition of cetuximab to radiotherapy and chemotherapy produced an ORR of 67%; EGFR gene copy number was not predictive of efficacy outcome in this trial<sup>411</sup>. A Phase 3 study of 938 patients with progressive NSCLC after platinum-based therapy concluded that the addition of cetuximab to chemotherapy was not recommended in this second-line setting<sup>412</sup>. Cetuximab is also being studied as part of a therapeutic regimen for patients with NSCLC with EGFR mutations who develop secondary resistance to erlotinib or gefitinib. A Phase 1b study combining afatinib and cetuximab for patients with either T790M-positive or T790M-negative tumors observed an overall ORR of 29%, and comparable response rates in both groups (32% T790M-positive vs. 25% T790M-negative)<sup>413</sup>. A Phase 1 study evaluating the combination erlotinib and cetuximab treatment for patients with NSCLC, including squamous tumors, regardless of EGFR status, as well as those who had progressed on prior erlotinib treatment, reported PRs in 10% (2/20) of patients and SDs lasting at least 6 months in 15% (3/20) of patients<sup>414</sup>; in addition, a retrospective analysis of this trial identified a patient with an exon 19 deletion and T790M who progressed rapidly on cetuximab and erlotinib<sup>415</sup>.

## Panitumumab

Assay findings association

**EGFR**  
amplification, L858R

### AREAS OF THERAPEUTIC USE

Panitumumab is a monoclonal antibody that targets EGFR. It is available in the EU to treat patients with RAS wild-type, metastatic colorectal cancer (CRC) combined with chemotherapy as first- or second-line therapy, or as monotherapy for patients who have progressed on prior chemotherapy. Please see the drug label for full prescribing information.

### GENE ASSOCIATION

For patients with metastatic CRC receiving cetuximab or panitumumab as mono- or combination therapy, increased EGFR copy number associated with improved OS (HR=0.62) in a meta-analysis, although increased

survival was not seen in populations that received first-line treatment with EGFR antibodies<sup>410</sup>.

### SUPPORTING DATA

In a Phase 2 trial for patients with advanced non-small cell lung cancer (NSCLC), the addition of panitumumab to paclitaxel/carboplatin did not result in improved clinical benefit<sup>416</sup>; a subsequent Phase 2 study investigating the addition of panitumumab to pemetrexed/cisplatin reported no benefit for patients with wildtype KRAS lung adenocarcinoma<sup>417</sup>. The combination of afatinib and panitumumab has been explored for 2 patients with EGFR T790M NSCLC, with 1 PR reported<sup>418</sup>.

**NOTE** Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies listed in this report may not be complete and exhaustive and the therapeutic agents are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type.

CLINICAL TRIALS

ORDERED TEST #

**NOTE** Clinical trials are ordered by gene and prioritized in the following descending order: Pediatric trial qualification → Geographical proximity → Trial phase → Trial verification within last 2 months. While every effort is made to ensure the accuracy of the information contained below, the

information available in the public domain is continually updated and should be investigated by the physician or research staff. The clinical trials listed in this report may not be complete and exhaustive or may include trials for which the patient does not meet the clinical trial

enrollment criteria. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](http://clinicaltrials.gov) or local registries in your region.

**GENE**  
**EGFR**

**ALTERATION**  
amplification, L858R

**RATIONALE**  
EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome

resistance to current agents include next-generation EGFR inhibitors and combination therapies.

**NCT04413201**

AFAMOSI: Efficacy and Safety of Afatinib Followed by Osimertinib Compared to Osimertinib in Patients With EGFRmutated/T790M Mutation Negative Nonsquamous NSCLC

**LOCATIONS:** München (Germany), Regensburg (Germany), Immenstadt (Germany), Löwenstein (Germany), Konstanz (Germany), Offenbach (Germany), Gießen (Germany), Hamm (Germany), Berlin (Germany), Bremen (Germany)

**PHASE 4**

**TARGETS**  
EGFR, ERBB4, ERBB2

**NCT04077463**

A Study of Lazertinib as Monotherapy or in Combination With JNJ-61186372 in Japanese Participants With Advanced Non-small Cell Lung Cancer

**LOCATIONS:** Gauting (Germany), Stuttgart (Germany), Frankfurt am Main (Germany), Monza (Italy), Milano (Italy), Halle (Saale) (Germany), Köln (Germany), Ravenna (Italy), Hemer (Germany), Essen (Germany)

**PHASE 1**

**TARGETS**  
EGFR, MET

**NCT04811001**

Best EGFR-TKI Sequence in NSCLC Harboring EGFR Mutations

**LOCATIONS:** Padova (Italy), Novara (Italy), Roma (Italy), Napoli (Italy)

**PHASE 2**

**TARGETS**  
EGFR, ERBB4, ERBB2

**NCT04487080**

A Study of Amivantamab and Lazertinib Combination Therapy Versus Osimertinib in Locally Advanced or Metastatic Non-Small Cell Lung Cancer

**LOCATIONS:** Monza (Italy), Milano (Italy), Rozzano (Italy), Szombathely (Hungary), Ravenna (Italy), Meldola (Italy), Orbassano (Italy), Szekesfehervar (Hungary), Charleroi (Belgium), Nijmegen (Netherlands)

**PHASE 3**

**TARGETS**  
MET, EGFR

**NCT03944772**

Phase 2 Platform Study in Patients With Advanced Non-Small Lung Cancer Who Progressed on First-Line Osimertinib Therapy (ORCHARD)

**LOCATIONS:** Maastricht (Netherlands), Nijmegen (Netherlands), Rotterdam (Netherlands), Amsterdam (Netherlands), Barcelona (Spain), Drammen (Norway), Oslo (Norway), Uppsala (Sweden), Madrid (Spain), A Coruña (Spain)

**PHASE 2**

**TARGETS**  
EGFR, PD-L1, RET, MET, ALK

CLINICAL TRIALS

ORDERED TEST #

<b>NCT04233021</b>	<b>PHASE 2</b>
Study of Osimertinib in Patients With a Lung Cancer With Brain or Leptomeningeal Metastases With EGFR Mutation	<b>TARGETS</b> EGFR
<b>LOCATIONS:</b> Strasbourg (France), Colmar (France), Besançon (France), Dijon (France), Grenoble (France), Villefranche-sur-Saône (France), Lyon (France), Pierre-Bénite (France), Amiens (France), Lille (France)	
<b>NCT02609776</b>	<b>PHASE 1</b>
A Dose Escalation Study of JNJ-61186372 in Participants With Advanced Non-Small Cell Lung Cancer	<b>TARGETS</b> MET, EGFR
<b>LOCATIONS:</b> Ravenna (Italy), Dijon (France), Lyon Cedex 8 (France), Villejuif Cedex (France), Paris (France), Marseille (France), Napoli (Italy), Sutton (United Kingdom), Saint-Herblain Cedex (France), Bordeaux (France)	
<b>NCT03810066</b>	<b>PHASE 2</b>
Exploring the Theragnostic Value of Osimertinib in EGFR-mutated Lung Cancer (THEROS)	<b>TARGETS</b> EGFR
<b>LOCATIONS:</b> Essen (Germany)	
<b>NCT03865511</b>	<b>PHASE 2</b>
MEchanisms of Resistance in EGFR Mutated Nonpretreated Advanced Lung Cancer Receiving OSimertib	<b>TARGETS</b> EGFR
<b>LOCATIONS:</b> Toulon (France), Le Mans (France), Cholet (France), Nantes (France)	
<b>NCT04721015</b>	<b>PHASE 1</b>
Study of Intravenous (IV) ABBV-637 Alone or in Combination With IV Docetaxel/Osimertinib to Assess Adverse Events and Change in Disease Activity in Adult Participants With Relapsed/Refractory (R/R) Solid Tumors	<b>TARGETS</b> EGFR
<b>LOCATIONS:</b> Marseille CEDEX 05 (France), Dijon (France), Toulouse (France), Bordeaux (France), Barcelona (Spain), Madrid (Spain), Majadahonda (Spain), Malaga (Spain), Ramat Gan (Israel), Haifa (Israel)	

ORDERED TEST #

**GENE**  
**MDM2**

**RATIONALE**  
Inhibitors of the MDM2-p53 interaction are being tested in clinical trials. Overexpression or

amplification of MDM2 may increase sensitivity to these agents, but more data are required.

**ALTERATION**  
amplification

**NCT04589845**

**PHASE 2**

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

**TARGETS**  
TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

**LOCATIONS:** München (Germany), Heilbronn (Germany), Würzburg (Germany), Zürich (Switzerland), Bellinzona (Switzerland), Basel (Switzerland), Brescia (Italy), Bern (Switzerland), Milano (Italy), Göttingen (Germany)

**NCT03449381**

**PHASE 1**

This Study Aims to Find the Best Dose of BI 907828 in Patients With Different Types of Advanced Cancer (Solid Tumors)

**TARGETS**  
MDM2

**LOCATIONS:** Tübingen (Germany), Köln (Germany), Berlin (Germany), Leuven (Belgium), Poznan (Poland), Warsaw (Poland), Barcelona (Spain), Ottawa (Canada), Connecticut, New York

**NCT03611868**

**PHASE 1/2**

A Study of APG-115 in Combination With Pembrolizumab in Patients With Metastatic Melanomas or Advanced Solid Tumors

**TARGETS**  
MDM2, PD-1

**LOCATIONS:** New York, Pennsylvania, District of Columbia, Virginia, Ohio, Tennessee, Missouri, Florida, Arkansas

**NCT04785196**

**PHASE 1/2**

APG-115 in Combination With PD-1 Inhibitor in Patients With Advanced Liposarcoma or Advanced Solid Tumors

**TARGETS**  
PD-1, MDM2

**LOCATIONS:** Shanghai (China), Guangzhou (China)

**NCT03725436**

**PHASE 1**

ALRN-6924 and Paclitaxel in Treating Patients With Advanced, Metastatic, or Unresectable Solid Tumors

**TARGETS**  
MDM2, MDM4

**LOCATIONS:** Texas



CLINICAL TRIALS

ORDERED TEST #

GENE  
**MTAP**

RATIONALE  
MTAP loss may predict sensitivity to MAT2A inhibitors.

ALTERATION  
loss

**NCT03435250**

PHASE 1

Study of AG-270 in Participants With Advanced Solid Tumors or Lymphoma With MTAP Loss

TARGETS  
MAT2A

LOCATIONS: Villejuif Cedex (France), Barcelona (Spain), Massachusetts, Connecticut, New York, Tennessee

SAMPLE

ORDERED TEST #

**APPENDIX** Variants of Unknown Significance

**NOTE** One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

**ARID1A**  
E1779G

**ASXL1**  
E865K

**ATR**  
I1062V

**CBL**  
R829W

**EPHB4**  
P79L

**ERBB2**  
R536Q

**FGF10**  
amplification

**MSH3**  
A60\_A62del

**NTRK2**  
P507K

**PTCH1**  
I1430V

**RICTOR**  
amplification

**SDHA**  
amplification

SAMPLE

ORDERED TEST #

APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

**DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS**

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMSB	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RADS1B	RADS1C	RADS1D	RADS2	RADS4L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENTSC (FAM46C)	TET2	TGFB2	TIPARP
TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL
WT1	XPO1	XRCC2	ZNF217	ZNF703				

**DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS**

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV1
ETV4	ETV5	ETV6	EWRS1	EZR	FGFR1	FGFR2	FGFR3	KIT
KMT2A (MLL)	MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA
RAF1	RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**
TMPPRSS2								

\*TERC is an NCRNA

\*\*Promoter region of TERT is interrogated


**ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER GENOMIC SIGNATURES**

- Homologous Recombination status
- Loss of Heterozygosity (LOH) score
- Microsatellite (MS) status
- Tumor Mutational Burden (TMB)

ORDERED TEST #

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplastraat 3, 2440 Geel, Belgium. 

**ABOUT FOUNDATIONONE CDX**

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details: [www.rochefoundationmedicine.com/f1cdxtech](http://www.rochefoundationmedicine.com/f1cdxtech).

**INTENDED USE**

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

**TEST PRINCIPLES**

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes. Using an Illumina® HiSeq platform, hybrid

capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

**THE REPORT**

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: The association of a therapy with a genomic alteration or signature does not necessarily indicate pharmacologic effectiveness (or lack thereof); no association of a therapy with a genomic alteration or signature does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness).

**Diagnostic Significance**

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

**Qualified Alteration Calls (Equivocal and Subclonal)**

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

**Ranking of Therapies and Clinical Trials**

*Ranking of Therapies in Summary Table*

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

*Ranking of Clinical Trials*

Pediatric trial qualification → Geographical proximity → Later trial phase.

**NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION**

Genomic signatures and gene alterations detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) ([www.nccn.org](http://www.nccn.org)). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each genomic signature or gene alteration. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to [NCCN.org](http://NCCN.org). NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

**Limitations**

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X,

© 2022 Foundation Medicine, Inc. All rights reserved.

ORDERED TEST #

APPENDIX

About FoundationOne®CDx

"MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.

2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation [https://www.accessdata.fda.gov/cdrh\\_docs/pdf17/P170019B.pdf](https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf). The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

3. Homologous Recombination status may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas (Coleman et al., 2017; 28916367). Samples with deleterious *BRCA1/2* alteration and/or Loss of Heterozygosity (LOH) score  $\geq 16\%$  will be reported as "HRD Positive" and samples with absence of these findings will be reported as "HRD Not Detected," agnostic of potential secondary *BRCA1/2* reversion alterations. Certain potentially deleterious missense or small in-frame deletions in *BRCA1/2* may not be classified as deleterious and, in the absence of an elevated LOH profile, samples with such mutations may be classified as "HRD Not Detected." A result of "HRD Not Detected" does not rule out the presence of a *BRCA1/2* alteration or an elevated LOH profile outside the assay performance characteristic limitations.

4. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be

Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.

5. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.

6. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.

7. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

\*Interquartile Range = 1<sup>st</sup> Quartile to 3<sup>rd</sup> Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of  $>10\%$ , and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene

ORDERED TEST #

APPENDIX

About FoundationOne®CDx

tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D (MLL2)*, *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

**LEVEL OF EVIDENCE NOT PROVIDED**

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

**NO GUARANTEE OF CLINICAL BENEFIT**

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

**NO GUARANTEE OF REIMBURSEMENT**

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

**TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN**

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such

as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

**SELECT ABBREVIATIONS**

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

**REFERENCE SEQUENCE INFORMATION**

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version 6.3.0

The median exon coverage for this sample is 935x

ORDERED TEST #

**APPENDIX**
**References**

1. Gatalica Z, et al. *Cancer Epidemiol. Biomarkers Prev.* (2014) PMID: 25392179
2. Kroemer G, et al. *Oncoimmunology* (2015) PMID: 26140250
3. Lal N, et al. *Oncoimmunology* (2015) PMID: 25949894
4. Le DT, et al. *N. Engl. J. Med.* (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Warth A, et al. *Virchows Arch.* (2016) PMID: 26637197
7. Ninomiya H, et al. *Br. J. Cancer* (2006) PMID: 16641899
8. Vanderwalde A, et al. *Cancer Med* (2018) PMID: 29436178
9. Zang YS, et al. *Cancer Med* (2019) PMID: 31270941
10. Dudley JC, et al. *Clin. Cancer Res.* (2016) PMID: 26880610
11. Takamochi K, et al. *Lung Cancer* (2017) PMID: 28676214
12. Pyllkkänen L, et al. *Environ. Mol. Mutagen.* (1997) PMID: 9329646
13. Gonzalez R, et al. *Ann. Oncol.* (2000) PMID: 11061602
14. Chen XQ, et al. *Nat. Med.* (1996) PMID: 8782463
15. Merlo A, et al. *Cancer Res.* (1994) PMID: 8174113
16. Kocarnik JM, et al. *Gastroenterol Rep (Oxf)* (2015) PMID: 26337942
17. You JF, et al. *Br. J. Cancer* (2010) PMID: 21081928
18. Bairwa NK, et al. *Methods Mol. Biol.* (2014) PMID: 24623249
19. Boland CR, et al. *Cancer Res.* (1998) PMID: 9823339
20. Pawlik TM, et al. *Dis. Markers* (2004) PMID: 15528785
21. Boland CR, et al. *Gastroenterology* (2010) PMID: 20420947
22. Samstein RM, et al. *Nat. Genet.* (2019) PMID: 30643254
23. Goodman AM, et al. *Mol. Cancer Ther.* (2017) PMID: 28835386
24. Goodman AM, et al. *Cancer Immunol Res* (2019) PMID: 31405947
25. Cristescu R, et al. *Science* (2018) PMID: 30309915
26. Ready N, et al. *J. Clin. Oncol.* (2019) PMID: 30785829
27. Hellmann MD, et al. *N. Engl. J. Med.* (2018) PMID: 29658845
28. Hellmann MD, et al. *Cancer Cell* (2018) PMID: 29657128
29. Hellmann MD, et al. *Cancer Cell* (2018) PMID: 29731394
30. Rozeman EA, et al. *Nat Med* (2021) PMID: 33558721
31. Sharma P, et al. *Cancer Cell* (2020) PMID: 32916128
32. Rizvi NA, et al. *Science* (2015) PMID: 25765070
33. Colli LM, et al. *Cancer Res.* (2016) PMID: 27197178
34. Wang VE, et al. *J Immunother Cancer* (2017) PMID: 28923100
35. Carbone DP, et al. *N. Engl. J. Med.* (2017) PMID: 28636851
36. Rizvi H, et al. *J. Clin. Oncol.* (2018) PMID: 29337640
37. Forde PM, et al. *N. Engl. J. Med.* (2018) PMID: 29658848
38. Miao D, et al. *Nat. Genet.* (2018) PMID: 30150660
39. Chae YK, et al. *Clin Lung Cancer* (2019) PMID: 30425022
40. Paz-Ares et al., 2019; ESMO Abstract LBA80
41. Hellmann MD, et al. *N. Engl. J. Med.* (2019) PMID: 31562796
42. Chalmers ZR, et al. *Genome Med* (2017) PMID: 28420421
43. Spigel et al., 2016; ASCO Abstract 9017
44. Xiao D, et al. *Oncotarget* (2016) PMID: 27009843
45. Shim HS, et al. *J Thorac Oncol* (2015) PMID: 26200269
46. Govindan R, et al. *Cell* (2012) PMID: 22980976
47. Ding L, et al. *Nature* (2008) PMID: 18948947
48. Imielinski M, et al. *Cell* (2012) PMID: 22980975
49. Kim Y, et al. *J. Clin. Oncol.* (2014) PMID: 24323028
50. Stein et al., 2019; DOI: 10.1200/PO.18.00376
51. Meng G, et al. *PLoS One* (2022) PMID: 35113949
52. Chen Y, et al. *J. Exp. Clin. Cancer Res.* (2019) PMID: 31088500
53. Yu H, et al. *J Thorac Oncol* (2019) PMID: 30253973
54. Pfeifer GP, et al. *Mutat. Res.* (2005) PMID: 15748635
55. Hill VK, et al. *Annu Rev Genomics Hum Genet* (2013) PMID: 23875803
56. Pfeifer GP, et al. *Oncogene* (2002) PMID: 12379884
57. Johnson BE, et al. *Science* (2014) PMID: 24336570
58. Choi S, et al. *Neuro-oncology* (2018) PMID: 29452419
59. Cancer Genome Atlas Research Network, et al. *Nature* (2013) PMID: 23636398
60. Briggs S, et al. *J. Pathol.* (2013) PMID: 23447401
61. Heitzer E, et al. *Curr. Opin. Genet. Dev.* (2014) PMID: 24583393
62. *Nature* (2012) PMID: 22810696
63. Roberts SA, et al. *Nat. Rev. Cancer* (2014) PMID: 25568919
64. Marabelle A, et al. *Lancet Oncol.* (2020) PMID: 32919526
65. Rossell R, et al. *Lancet Oncol.* (2012) PMID: 22285168
66. Douillard JY, et al. *Br. J. Cancer* (2014) PMID: 24263064
67. Sequist LV, et al. *J. Clin. Oncol.* (2013) PMID: 23816960
68. Mok TS, et al. *J. Clin. Oncol.* (2018) PMID: 29864379
69. Jänne PA, et al. *N. Engl. J. Med.* (2015) PMID: 25923549
70. Hong MH, et al. *Cancer* (2020) PMID: 32749686
71. Kim HS, et al. *Oncotarget* (2015) PMID: 26462025
72. Kim HS, et al. *Clin. Cancer Res.* (2015) PMID: 25424851
73. Mondal G, et al. *Acta Neuropathol* (2020) PMID: 32303840
74. Cavallieri S, et al. *Eur. J. Cancer* (2018) PMID: 29734047
75. Chi AS, et al. *JCO Precis Oncol* (2020) PMID: 32923886
76. Pirker R, et al. *Lancet Oncol.* (2012) PMID: 22056021
77. Herbst RS, et al. *Lancet Oncol.* (2018) PMID: 29169877
78. Paz-Ares L, et al. *Ann. Oncol.* (2016) PMID: 27207107
79. Zhang X, et al. *J. Investig. Med.* (2017) PMID: 27664271
80. Dahabreh IJ, et al. *Ann. Oncol.* (2011) PMID: 20826716
81. Dahabreh IJ, et al. *Clin. Cancer Res.* (2010) PMID: 20028749
82. Carlson JJ, et al. *J Cancer Res Clin Oncol* (2009) PMID: 19430813
83. Fukuoka M, et al. *J. Clin. Oncol.* (2011) PMID: 21670455
84. Cappuzzo F, et al. *J Thorac Oncol* (2015) PMID: 25514804
85. De Grève J, et al. *Lung Cancer* (2015) PMID: 25682316
86. Crinò L, et al. *J Clin Oncol* (2008) PMID: 18779612
87. Kim ES, et al. *Lancet* (2008) PMID: 19027483
88. Soh J, et al. *Int J Cancer* (2007) PMID: 17487844
89. Goss GD, et al. *JAMA Oncol* (2018) PMID: 29902295
90. Wang F, et al. *J Transl Med* (2013) PMID: 23557218
91. Leighl et al., 2021; ESMO Abstract 1192MO
92. Cho et al., 2020; ESMO Abstract 12580
93. Bauml et al., 2021; ASCO Abstract 9006
94. Shu et al., 2021; ESMO Abstract 1193MO
95. Jänne PA, et al. *Cancer Discov* (2021) PMID: 34548309
96. Ahn MJ, et al. *Lancet Respir Med* (2017) PMID: 29056570
97. Yang Z, et al. *Sci Transl Med* (2016) PMID: 27928026
98. Ahn MJ, et al. *Lancet Oncol* (2019) PMID: 31587882
99. Lin L, et al. *Lung Cancer* (2022) PMID: 35248866
100. Reck M, et al. *Lancet Respir Med* (2019) PMID: 30922878
101. Socinski MA, et al. *J Thorac Oncol* (2021) PMID: 34311108
102. Socinski MA, et al. *N. Engl. J. Med.* (2018) PMID: 29863955
103. Lu et al., 2021; ESMO Abstract VPP9-2021
104. Vallee A, et al. *Int. J. Oncol.* (2013) PMID: 23934203
105. *Nature* (2014) PMID: 25079552
106. *Nature* (2012) PMID: 22960745
107. Park S, et al. *Histol. Histopathol.* (2012) PMID: 22207554
108. Liang Z, et al. *BMC Cancer* (2010) PMID: 20637128
109. Grob TJ, et al. *Lung Cancer* (2013) PMID: 23238037
110. Watzka SB, et al. *Eur J Cardiothorac Surg* (2010) PMID: 20353893
111. Dobashi Y, et al. *Hum. Pathol.* (2011) PMID: 21040950
112. Ludovini V, et al. *Cancer Chemother. Pharmacol.* (2013) PMID: 23314677
113. Skrzypski M, et al. *Clin Lung Cancer* (2013) PMID: 23870818
114. Kim SH, et al. *Histol. Histopathol.* (2012) PMID: 22419022
115. Lee JS, et al. *Ann. Surg. Oncol.* (2013) PMID: 23525704
116. Oakley GJ, et al. *J Thorac Oncol* (2011) PMID: 21587084
117. Marks JL, et al. *J Thorac Oncol* (2008) PMID: 18303429
118. Izar B, et al. *Ann. Thorac. Surg.* (2013) PMID: 23932319
119. Ciardiello F, et al. *N. Engl. J. Med.* (2008) PMID: 18337605
120. Bhargava R, et al. *Mod. Pathol.* (2005) PMID: 15920544
121. Yang YL, et al. *Chin. Med. J.* (2012) PMID: 22490401
122. Lynch TJ, et al. *N. Engl. J. Med.* (2004) PMID: 15118073
123. Paez JG, et al. *Science* (2004) PMID: 15118125
124. Pao W, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2004) PMID: 15329413
125. Yang JC, et al. *Lancet Oncol.* (2015) PMID: 25589191
126. Cheok CF, et al. *Nat Rev Clin Oncol* (2011) PMID: 20975744
127. Ohnstad HO, et al. *Cancer* (2013) PMID: 23165797
128. Gamble LD, et al. *Oncogene* (2012) PMID: 21725357
129. Zhang et al., 2019; ASCO Abstract 3124
130. Rasco et al., 2019; ASCO Abstract 3126
131. Tolcher et al., 2021; ASCO Abstract 2506
132. Martinelli et al., 2016; EHA21 Abstract S504
133. Daver et al., 2018; ASH Abstract 767
134. Mascarenhas et al., 2019; ASH Abstract 134
135. Shustov et al., 2018; ASH Abstract 1623
136. Sallman et al., 2018; ASH Abstract 4066
137. Meric-Bernstam et al., 2017; ASCO Abstract 2505
138. Higashiyama M, et al. *Br. J. Cancer* (1997) PMID: 9155050
139. Marchetti A, et al. *Diagn. Mol. Pathol.* (1995) PMID: 7551299
140. Dworakowska D, et al. *Lung Cancer* (2004) PMID: 15165086
141. Onel K, et al. *Mol. Cancer Res.* (2004) PMID: 14757840
142. Ren YW, et al. *Asian Pac. J. Cancer Prev.* (2013) PMID: 24175836
143. Sdek P, et al. *Mol. Cell* (2005) PMID: 16337594
144. Brady M, et al. *Mol. Cell. Biol.* (2005) PMID: 15632057
145. Li M, et al. *Mol. Cell* (2004) PMID: 15053880
146. Brown CJ, et al. *Nat. Rev. Cancer* (2009) PMID: 19935675
147. Cordon-Cardo C, et al. *Cancer Res.* (1994) PMID: 8306343
148. Beroukhim R, et al. *Nature* (2010) PMID: 20164920
149. Kato S, et al. *Clin. Cancer Res.* (2017) PMID: 28351930
150. Singavi et al., 2017; ESMO Abstract 1140PD
151. Kalev P, et al. *Cancer Cell* (2021) PMID: 33450196
152. Marjon K, et al. *Cell Rep* (2016) PMID: 27068473
153. Mavrakis KJ, et al. *Science* (2016) PMID: 26912361
154. Kryukov GV, et al. *Science* (2016) PMID: 26912360
155. Heist et al., 2019; AACR-NCI-EORTC Abstract B116
156. Guccione E, et al. *Nat. Rev. Mol. Cell Biol.* (2019) PMID: 31350521
157. Fedoriw A, et al. *Cancer Cell* (2019) PMID: 31257072
158. Srour N, et al. *Cancer Cell* (2019) PMID: 31287990
159. Gao G, et al. *Nucleic Acids Res.* (2019) PMID: 30916320

ORDERED TEST #

**APPENDIX**
**References**

160. Smith CR, et al. *J Med Chem* (2022) PMID: 35041419
161. Hansen LJ, et al. *Cancer Res.* (2019) PMID: 31040154
162. Tang B, et al. *Cancer Res.* (2018) PMID: 29844120
163. Munshi PN, et al. *Oncologist* (2014) PMID: 24928612
164. de Oliveira SF, et al. *PLoS ONE* (2016) PMID: 26751376
165. Lubin M, et al. *PLoS ONE* (2009) PMID: 19478948
166. Tang B, et al. *Cancer Biol. Ther.* (2012) PMID: 22825330
167. Collins CC, et al. *Mol. Cancer Ther.* (2012) PMID: 22252602
168. Bertino JR, et al. *Cancer Biol. Ther.* (2011) PMID: 21301207
169. Coulthard SA, et al. *Mol. Cancer Ther.* (2011) PMID: 21282358
170. Miyazaki S, et al. *Int. J. Oncol.* (2007) PMID: 17912432
171. Efferth T, et al. *Blood Cells Mol. Dis.* ( ) PMID: 11987241
172. Kindler HL, et al. *Invest New Drugs* (2009) PMID: 18618081
173. Alhalabi O, et al. *Nat Commun* (2022) PMID: 35379845
174. Wei R, et al. *Sci Rep* (2016) PMID: 27929028
175. Zhao M, et al. *BMC Genomics* (2016) PMID: 27556634
176. Kirovski G, et al. *Am. J. Pathol.* (2011) PMID: 21356366
177. Huang HY, et al. *Clin. Cancer Res.* (2009) PMID: 19887491
178. Marcé S, et al. *Clin. Cancer Res.* (2006) PMID: 16778103
179. Meyer S, et al. *Exp. Dermatol.* (2010) PMID: 20500769
180. Wild PJ, et al. *Arch Dermatol* (2006) PMID: 16618867
181. Kim J, et al. *Genes Chromosomes Cancer* (2011) PMID: 21412930
182. Li CF, et al. *Oncotarget* (2014) PMID: 25426549
183. He HL, et al. *Medicine (Baltimore)* (2015) PMID: 26656376
184. Su CY, et al. *Eur J Surg Oncol* (2014) PMID: 24969958
185. Mirebeau D, et al. *Haematologica* (2006) PMID: 16818274
186. Becker AP, et al. *Pathobiology* (2015) PMID: 26088413
187. Snezhkina AV, et al. *Oxid Med Cell Longev* (2016) PMID: 27433286
188. Bistulfi G, et al. *Oncotarget* (2016) PMID: 26910893
189. Antonopoulou K, et al. *J. Invest. Dermatol.* (2015) PMID: 25407435
190. Maccioni L, et al. *BMC Cancer* (2013) PMID: 23816148
191. Hyland PL, et al. *Int J Epidemiol* (2016) PMID: 26635288
192. Lin X, et al. *Cancer Sci.* (2017) PMID: 27960044
193. Zhi L, et al. *J Cancer* (2016) PMID: 27994653
194. Gu F, et al. *Br. J. Cancer* (2013) PMID: 23361049
195. Limm K, et al. *PLoS ONE* (2016) PMID: 27479139
196. Tang B, et al. *G3 (Bethesda)* (2014) PMID: 25387827
197. Limm K, et al. *Eur. J. Cancer* (2013) PMID: 23265702
198. Stevens AP, et al. *J. Cell. Biochem.* (2009) PMID: 19097084
199. Limm K, et al. *Eur. J. Cancer* (2014) PMID: 25087184
200. Elvin JA, et al. *Oncologist* (2017) PMID: 28283584
201. Gao J, et al. *Curr Oncol* (2015) PMID: 26715889
202. Gopalan et al., 2014; *ASCO Abstract* 8077
203. Peguero et al., 2016; *ASCO Abstract* 2528
204. Konecny et al., 2016; *ASCO Abstract* 5557
205. DeMichele A, et al. *Clin. Cancer Res.* (2015) PMID: 25501126
206. Finn RS, et al. *Lancet Oncol.* (2015) PMID: 25524798
207. Infante JR, et al. *Clin. Cancer Res.* (2016) PMID: 27542767
208. Johnson DB, et al. *Oncologist* (2014) PMID: 24797823
209. Van Maerken T, et al. *Mol. Cancer Ther.* (2011) PMID: 21460101
210. Konecny GE, et al. *Clin. Cancer Res.* (2011) PMID: 21278246
211. Katsumi Y, et al. *Biochem. Biophys. Res. Commun.* (2011) PMID: 21871868
212. Cen L, et al. *Neuro-oncology* (2012) PMID: 22711607
213. Logan JE, et al. *Anticancer Res.* (2013) PMID: 23898052
214. Shapiro N, et al., 2013; *ASCO Abstract* 2500
215. Flaherty KT, et al. *Clin. Cancer Res.* (2012) PMID: 22090362
216. Dickson MA, et al. *J. Clin. Oncol.* (2013) PMID: 23569312
217. Doxtader EE, et al. *Hum. Pathol.* (2012) PMID: 21840041
218. Gazzeri S, et al. *Oncogene* (1998) PMID: 9484839
219. Kratzke RA, et al. *Cancer Res.* (1996) PMID: 8758904
220. Lee JU, et al. *Tuberc Respir Dis (Seoul)* (2012) PMID: 23101020
221. Cortot AB, et al. *Clin Lung Cancer* (2014) PMID: 24169260
222. Mounawar M, et al. *Cancer Res.* (2007) PMID: 17575133
223. Zhao Y, et al. *Clin Lung Cancer* (2011) PMID: 21889114
224. Kawabuchi B, et al. *Int. J. Cancer* (1999) PMID: 9988232
225. Xing XB, et al. *PLoS ONE* (2013) PMID: 23805242
226. Lou-Qian Z, et al. *PLoS ONE* (2013) PMID: 23372805
227. Quelle DE, et al. *Cell* (1995) PMID: 8521522
228. *Mutat. Res.* (2005) PMID: 15878778
229. *Oncogene* (1999) PMID: 10498883
230. Sherr CJ, et al. *Cold Spring Harb. Symp. Quant. Biol.* (2005) PMID: 16869746
231. Ozenne P, et al. *Int. J. Cancer* (2010) PMID: 20549699
232. Ruas M, et al. *Oncogene* (1999) PMID: 10498896
233. Jones R, et al. *Cancer Res.* (2007) PMID: 17909018
234. Haferkamp S, et al. *Aging Cell* (2008) PMID: 18843795
235. Huot TJ, et al. *Mol. Cell. Biol.* (2002) PMID: 12417717
236. Rizo H, et al. *J. Biol. Chem.* (2001) PMID: 11518711
237. Gombart AF, et al. *Leukemia* (1997) PMID: 9324288
238. Yang R, et al. *Cancer Res.* (1995) PMID: 7780957
239. Parry D, et al. *Mol. Cell. Biol.* (1996) PMID: 8668202
240. Greenblatt MS, et al. *Oncogene* (2003) PMID: 12606942
241. Yarbrough WG, et al. *J. Natl. Cancer Inst.* (1999) PMID: 10491434
242. Poi MJ, et al. *Mol. Carcinog.* (2001) PMID: 11255261
243. Byeon IJ, et al. *Mol. Cell* (1998) PMID: 9660926
244. Kannengiesser C, et al. *Hum. Mutat.* (2009) PMID: 19260062
245. Lal G, et al. *Genes Chromosomes Cancer* (2000) PMID: 10719365
246. Koh J, et al. *Nature* (1995) PMID: 7777061
247. McKenzie HA, et al. *Hum. Mutat.* (2010) PMID: 20340136
248. Miller PJ, et al. *Hum. Mutat.* (2011) PMID: 21462282
249. Kutscher CL, et al. *Physiol. Behav.* (1977) PMID: 905385
250. Scaini MC, et al. *Hum. Mutat.* (2014) PMID: 24659262
251. Jenkins NC, et al. *J. Invest. Dermatol.* (2013) PMID: 23190892
252. Walker GJ, et al. *Int. J. Cancer* (1999) PMID: 10389768
253. Rutter JL, et al. *Oncogene* (2003) PMID: 12853981
254. Itahana K, et al. *Cancer Cell* (2008) PMID: 18538737
255. Zhang Y, et al. *Mol. Cell* (1999) PMID: 10360174
256. Zhang Y, et al. *Cell* (1998) PMID: 9529249
257. Jafri M, et al. *Cancer Discov* (2015) PMID: 25873077
258. Whelan AJ, et al. *N Engl J Med* (1995) PMID: 7666917
259. *Adv Exp Med Biol* (2010) PMID: 20687502
260. Hogg D, et al. *J Cutan Med Surg* (1998) PMID: 9479083
261. De Unamuno B, et al. *Melanoma Res* (2018) PMID: 29543703
262. Soura E, et al. *J Am Acad Dermatol* (2016) PMID: 26892650
263. Huerta C, et al. *Acta Derm Venereol* (2018) PMID: 29405243
264. Kaufman DK, et al. *Neurology* (1993) PMID: 8414022
265. Bahau M, et al. *Cancer Res* (1998) PMID: 9622062
266. Chan AK, et al. *Clin Neuropathol* ( ) PMID: 28699883
267. Beuselinck B, et al. *Acta Oncol* (2018) PMID: 29095068
268. Song Y, et al. *Chin. Med. J.* (2015) PMID: 26228213
269. Dornbusch J, et al. *PLoS ONE* (2013) PMID: 24086736
270. Terakawa T, et al. *Urol. Oncol.* (2013) PMID: 21478036
271. You D, et al. *World J Urol* (2015) PMID: 24710685
272. Silva E, et al. *Breast J* ( ) PMID: 25639617
273. Baumgarten P, et al. *Neuro-oncology* (2016) PMID: 26627848
274. Sathornsumetee S, et al. *J. Clin. Oncol.* (2008) PMID: 18182667
275. Olafson LR, et al. *J Clin Neurosci* (2019) PMID: 31582283
276. Duda DG, et al. *Oncologist* (2010) PMID: 20484123
277. Stremtizer S, et al. *Mol. Cancer Ther.* (2016) PMID: 27535973
278. Weickhardt AJ, et al. *Br. J. Cancer* (2015) PMID: 26125443
279. Kopetz S, et al. *J. Clin. Oncol.* (2010) PMID: 20008624
280. Miles DW, et al. *Br. J. Cancer* (2013) PMID: 23422754
281. Fountzilias G, et al. *Anticancer Res.* (2011) PMID: 21868552
282. Gianni L, et al. *J. Clin. Oncol.* (2013) PMID: 23569311
283. Sánchez-Rovira P, et al. *Clin Transl Oncol* (2013) PMID: 23397155
284. Cameron D, et al. *Lancet Oncol.* (2013) PMID: 23932548
285. Mok T, et al. *J Thorac Oncol* (2014) PMID: 24807156
286. An SJ, et al. *Cancer Gene Ther.* (2014) PMID: 24577128
287. Bais C, et al. *J. Natl. Cancer Inst.* (2017) PMID: 29059426
288. Cohen EE, et al. *Lancet Oncol.* (2009) PMID: 19201650
289. Van Cutsem E, et al. *J. Clin. Oncol.* (2012) PMID: 22565005
290. Lee EQ, et al. *Clin. Cancer Res.* (2018) PMID: 29941486
291. Xu L, et al. *Cancer Res.* (2009) PMID: 19826039
292. Heist RS, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2015) PMID: 25605928
293. Zietsch J, et al. *PLoS ONE* (2010) PMID: 20686603
294. Holtkamp N, et al. *Neuro-oncology* (2007) PMID: 17504929
295. Ramos AH, et al. *Cancer Biol. Ther.* (2009) PMID: 19755855
296. Seto T, et al. *Lung Cancer* (2006) PMID: 16697074
297. Yang F, et al. *Cancer Res.* (2011) PMID: 21724587
298. *Biol. Pharm. Bull.* (2011) PMID: 22130231
299. Tate JG, et al. *Nucleic Acids Res.* (2019) PMID: 30371878
300. Hegde M, et al. *Genet. Med.* (2014) PMID: 24310308
301. Aretz S, et al. *Eur. J. Hum. Genet.* (2013) PMID: 22872101
302. Win AK, et al. *Gastroenterology* (2014) PMID: 24444654
303. Lubbe SJ, et al. *J. Clin. Oncol.* (2009) PMID: 19620482
304. Jones N, et al. *Gastroenterology* (2009) PMID: 19394335
305. Nielsen M, et al. *J. Natl. Cancer Inst.* (2010) PMID: 21044966
306. David SS, et al. *Nature* (2007) PMID: 17581577
307. Molatore S, et al. *Hum. Mutat.* (2010) PMID: 19953527
308. Kundu S, et al. *DNA Repair (Amst.)* (2009) PMID: 19836313
309. D'Agostino VG, et al. *DNA Repair (Amst.)* (2010) PMID: 20418187
310. Ali M, et al. *Gastroenterology* (2008) PMID: 18534194
311. Landrum MJ, et al. *Nucleic Acids Res.* (2018) PMID: 29165669
312. Sampson JR, et al. *Lancet* (2003) PMID: 12853198
313. Sieber OM, et al. *N. Engl. J. Med.* (2003) PMID: 12606733
314. Al-Tassan N, et al. *Nat. Genet.* (2002) PMID: 11818965
315. Rennert G, et al. *Cancer* (2012) PMID: 21952991
316. Zhang Y, et al. *Cancer Epidemiol. Biomarkers Prev.*



ORDERED TEST #

**APPENDIX**
**References**

- (2006) pmid: 16492928
317. von der Thüsen JH, et al. *J. Clin. Oncol.* (2011) pmid: 21189386
318. Casper M, et al. *Fam. Cancer* (2014) pmid: 24420788
319. Smith LM, et al. *Pancreatology* (2009) pmid: 20110747
320. Zehir A, et al. *Nat. Med.* (2017) pmid: 28481359
321. Nguyen B, et al. *Cell* (2022) pmid: 35120664
322. Yin LL, et al. *Oncol Lett* (2018) pmid: 30405763
323. Li Z, et al. *Cell Cycle* (2020) pmid: 33064970
324. Witkiewicz AK, et al. *Nat Commun* (2015) pmid: 2585536
325. Zhao Z, et al. *Eur Rev Med Pharmacol Sci* (2020) pmid: 32572914
326. Xiao W, et al. *Am J Cancer Res* (2021) pmid: 33520366
327. Sun X, et al. *Int J Oncol* (2019) pmid: 30483773
328. Wang Y, et al. *EMBO Mol Med* (2013) pmid: 24000153
329. Inoue A, et al. *FEBS Lett.* (2014) pmid: 24530524
330. Johnston JJ, et al. *Am. J. Hum. Genet.* (2010) pmid: 20451169
331. Gripp KW, et al. *Am. J. Med. Genet. A* (2011) pmid: 21910224
332. Wu YL, et al. *Lancet Oncol.* (2014) pmid: 24439929
333. Passaro et al., 2019; ELCC Abstract 1150
334. Audet et al., 2013; ASCO Abstract 6041
335. Lau SC, et al. *Clin Lung Cancer* (2019) pmid: 31178389
336. Paz-Ares L, et al. *Ann. Oncol.* (2017) pmid: 28426106
337. Thongprasert S, et al. *Lung Cancer Manag* (2019) pmid: 31807143
338. Januszewski et al., 2018; IASLC WCLC Abstract P1.13-17
339. Suzuki et al., 2018; IASLC WCLC Abstract P1.01-92
340. Chang et al., 2018; IASLC WCLC Abstract P1.01-11
341. Llinás-Quintero N, et al. *Case Rep Oncol Med* (2019) pmid: 31637072
342. Miller VA, et al. *Lancet Oncol.* (2012) pmid: 22452896
343. Chen X, et al. *Lung Cancer* (2013) pmid: 23664448
344. Katakami N, et al. *J. Clin. Oncol.* (2013) pmid: 23816963
345. Landi L, et al. *Clin Lung Cancer* (2014) pmid: 25242668
346. Yang JC, et al. *Lancet Oncol.* (2015) pmid: 26051236
347. Horn L, et al. *Lung Cancer* (2017) pmid: 29110849
348. Yamamoto N, et al. *Adv Ther* (2020) pmid: 31863283
349. Soria JC, et al. *Lancet Oncol.* (2015) pmid: 26156651
350. Dziadziuszko R, et al. *J Thorac Oncol* (2019) pmid: 30825613
351. Lai WV, et al. *Eur. J. Cancer* (2019) pmid: 30685684
352. Greulich H, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2012) pmid: 22908275
353. Gow CH, et al. *J Thorac Oncol* (2015) pmid: 26134234
354. Mazières J, et al. *Ann. Oncol.* (2016) pmid: 26598547
355. Mazières J, et al. *J. Clin. Oncol.* (2013) pmid: 23610105
356. De Grève J, et al. *Lung Cancer* (2012) pmid: 22325357
357. Li BT, et al. *Lung Cancer* (2015) pmid: 26559459
358. Costa DB, et al. *J Thorac Oncol* (2016) pmid: 26964772
359. Yuan B, et al. *Front Oncol* (2020) pmid: 32477948
360. Fang W, et al. *Oncologist* (2019) pmid: 31748336
361. Schuler M, et al. *Ann. Oncol.* (2016) pmid: 26646759
362. Wu YL, et al. *Lancet Oncol.* (2017) pmid: 28958502
363. Opsomer RJ, et al. *Acta Urol Belg* (1985) pmid: 2986437
364. Wu et al., 2018; WCLC abstract MA26.11
365. Ramalingam SS, et al. *Ann. Oncol.* (2016) pmid: 26768165
366. Yu HA, et al. *Lung Cancer* (2017) pmid: 29191595
367. Reckamp KL, et al. *Cancer* (2014) pmid: 24501009
368. Jänne PA, et al. *Clin. Cancer Res.* (2011) pmid: 21220471
369. van Geel RMJM, et al. *Br. J. Cancer* (2020) pmid: 32147669
370. Jänne PA, et al. *J Thorac Oncol* (2016) pmid: 26899759
371. Cappuzzo F, et al. *Lancet Oncol.* (2010) pmid: 20493771
372. Zhong WZ, et al. *J. Clin. Oncol.* (2019) pmid: 31194613
373. Petrelli F, et al. *Clin Lung Cancer* (2012) pmid: 22056888
374. Petty RD, et al. *J. Clin. Oncol.* (2017) pmid: 28537764
375. Philip PA, et al. *J. Clin. Oncol.* (2006) pmid: 16809731
376. Xie C, et al. *Br J Cancer* (2020) pmid: 32958820
377. Luo H, et al. *JAMA Netw Open* (2020) pmid: 33026449
378. Lee J, et al. *Lancet Oncol.* (2012) pmid: 22192731
379. Leon et al., 2014; doi.org/10.1093/annonc/mdu349.52
380. Lee CK, et al. *J. Natl. Cancer Inst.* (2017) pmid: 28376144
381. Yang JJ, et al. *Br. J. Cancer* (2017) pmid: 28103612
382. Zhou Q, et al. *Cancer Cell* (2021) pmid: 34388377
383. Kawashima Y, et al. *Lancet Respir Med* (2022) pmid: 34454653
384. Saito H, et al. *Lancet Oncol* (2019) pmid: 30975627
385. Piccirillo et al., 2021; ESMO Abstract 12070
386. Faehling M, et al. *J Cancer Res Clin Oncol* (2018) pmid: 29687154
387. Nakagawa K, et al. *Lancet Oncol.* (2019) pmid: 31591063
388. Han JY, et al. *J. Clin. Oncol.* (2012) pmid: 22370314
389. Maemondo M, et al. *N. Engl. J. Med.* (2010) pmid: 20573926
390. Mitsudomi T, et al. *Lancet Oncol.* (2010) pmid: 20022809
391. Mok TS, et al. *N. Engl. J. Med.* (2009) pmid: 19692680
392. Qi WX, et al. *Curr Med Res Opin* (2015) pmid: 25329826
393. Zhao H, et al. *J Thorac Oncol* (2015) pmid: 25546556
394. Wang J, et al. *Int. J. Cancer* (2019) pmid: 30255937
395. Baik CS, et al. *J Thorac Oncol* (2015) pmid: 26398831
396. Dutton Y, et al. *Lancet Oncol.* (2014) pmid: 24950987
397. Yoshioka H, et al. *Ann. Oncol.* (2019) pmid: 31553438
398. Sutiman N, et al. *J Thorac Oncol* (2017) pmid: 27908825
399. Noronha V, et al. *J. Clin. Oncol.* (2019) pmid: 31411950
400. Hosomi Y, et al. *J. Clin. Oncol.* (2020) pmid: 31682542
401. Creelan BC, et al. *Br J Cancer* (2021) pmid: 33012782
402. Soria JC, et al. *N. Engl. J. Med.* (2018) pmid: 29151359
403. Alanazi A, et al. *Lung Cancer Manag* (2020) pmid: 33318755
404. Kim et al., 2021; DOI: 10.1200/PO.20.00296
405. Ramalingam SS, et al. *N. Engl. J. Med.* (2019) pmid: 31751012
406. Herbst et al., 2020; ASCO Abstract LBA5
407. Kenmotsu et al., 2021; ESMO Abstract LBA44
408. Soo et al., 2021; ESMO Abstract VP3-2021
409. Oxnard GR, et al. *Ann. Oncol.* (2020) pmid: 32139298
410. Jiang Z, et al. *PLoS ONE* (2013) pmid: 23441167
411. Ramalingam SS, et al. *Lung Cancer* (2013) pmid: 23849982
412. Kim ES, et al. *Lancet Oncol.* (2013) pmid: 24231627
413. Janjigian YY, et al. *Cancer Discov* (2014) pmid: 25074459
414. Wheler JJ, et al. *Mol. Cancer Ther.* (2013) pmid: 23963360
415. Tsigelny IF, et al. *Oncotarget* (2015) pmid: 25760241
416. Crawford J, et al. *J Thorac Oncol* (2013) pmid: 24389433
417. Schuette W, et al. *Clin Lung Cancer* (2015) pmid: 26094080
418. Castellanos EH, et al. *Clin Lung Cancer* (2015) pmid: 25842367